

Table 1 Suppl. List of primers (LP - left primer, RP - right primer, LB - left border primer, F - forward, R - reverse, RT-qPCR - reverse transcription quantitative PCR).

Targets	Primers (5'→3')	Experiment	Reference
<i>FLOT1</i>	LP: GGGACAAAGGAGTTTAAGAAGG RP: GTTCCGCACCACGTAGAGTAC	genotyping, RT-PCR	this study
<i>FLOT2</i>	LP: TACCACTCCCACTAGCACCAC RP: TGTGAAGGTGTTATCGAGGG	genotyping, RT-PCR	this study
<i>FLOT3</i>	LP: TCCCTTCTCCTAGCCTTTGAG RP: TGTAATAAACC GCGTTTCAATG	genotyping	this study
<i>FLOT3</i>	LP: GGTGTTTCCATGGCAGTCTT RP: GCTGATCTTAGGCTGCAGGT	RT-PCR	this study
T-DNA FLAG line	LB: CGTGTGCCAGGTGCCACGGAATAGT	genotyping	this study
T-DNA SALK lines	LB: ATTTTGCCGATTTTCGGAAC	genotyping	this study
<i>Actin2</i>	F: CCGCTCTTTCTTTCCAAGC R: CCGGTACCATTGTACACAC	RT-PCR	this study
<i>AtFLOT1</i>	F: ATGAACGCTTTGACTCGAAC R: GGCTTGCTTTTGTTCCTCGTA	RT-qPCR	Li <i>et al.</i> 2012
<i>AtFLOT2</i>	F: ACTTGCAGCCCAAGATTAGC R: CTCCCACTAGCACCACCAAT	RT-qPCR	this study
<i>AtFLOT3</i>	F: AGTCGCTAAAGCATCGCAGT R: TGCAAGCTTGATGTCTGTGA	RT-qPCR	this study
<i>TIP41</i>	F: GTGAAAACCTGTTGGAGAGAAGCAA R: TCAACTGGATACCCTTTTCGCA	RT-qPCR	Czechowski <i>et al.</i> 2005

Czechowski, T., Stitt, M., Altmann, T., Udvardi, M.K., Scheible, W.R.: Genome-wide identification and testing of superior reference genes for transcript normalization in *Arabidopsis*. - *Plant Physiol.* **139**: 5-17, 2005.

Li, R., Liu, P., Wan, Y., Chen, T., Wang, Q., Mettbach, U., Baluska, F., Samaj, J., Fang, X., Lucas, W.J., Lin, J.: A membrane microdomain-associated protein, *Arabidopsis* Flot1, is involved in a clathrin-independent endocytic pathway and is required for seedling development. - *Plant Cell* **24**: 2105-2122, 2012.

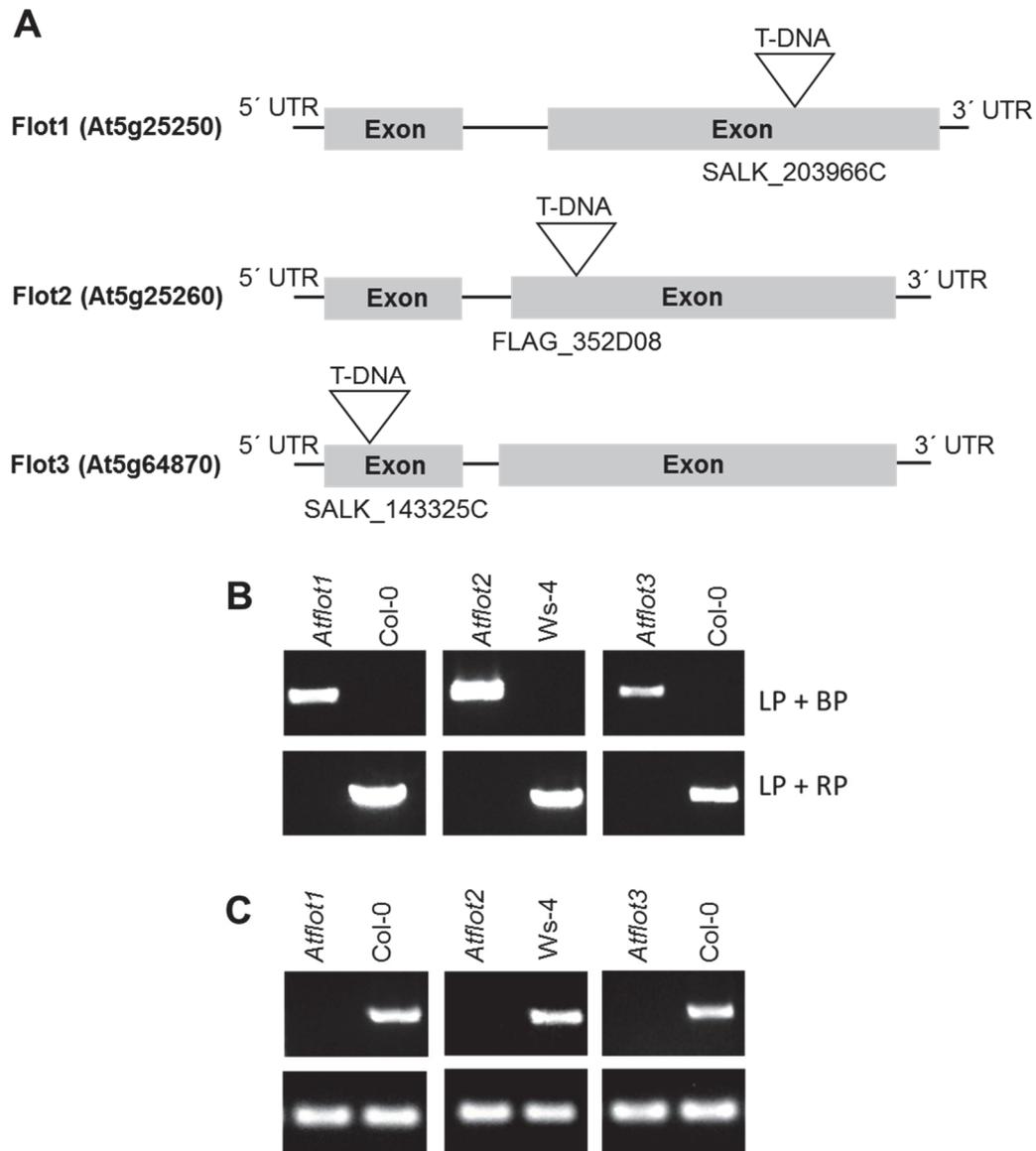


Fig. 1 Suppl. Characterization of T-DNA mutants. *A* - Schematic overview of T-DNA insertion in particular AtFLOT knock-out mutants. *B* - Genomic PCR analysis to confirm the integration of T-DNA in *Atf1*, *Atf2*, and *Atf3* (*upper panels* - PCR products of primers used for amplification of T-DNA insertion allele, *lower panels* - PCR products of primers used for amplification of wild-type allele, LP - left primer, RP - right primer, BP - T-DNA border primer). *C* - RT-PCR analysis of *flot1*, *flot2*, and *flot3* specific transcripts (*upper panels* - RT-PCR with gene specific primers, *lower panels* - *Actin2* used as an internal control).

DNA was isolated from 3-week-old *Atf1* (SALK_203966), *Atf2* (FLAG_352D08), and *Atf3* (SALK_143325C) plants. T-DNA insertion was confirmed by PCR using primers listed in Table 1 Suppl. For RT-PCR analysis the leaf samples were instantly frozen in liquid nitrogen. RNA was isolated using a *Spectrum Plant Total RNA kit* (*Sigma-Aldrich*, St. Louis USA), *Turbo DNA-free kit* (*Applied Biosystems*, Foster City, USA) was used for DNA removal and *Transcriptor High Fidelity cDNA synthesis kit* (*Roche*, Basel, Switzerland) was used for cDNA synthesis. The reverse transcription reaction was primed with anchored-oligo(DT)18 primer. Primers used for RT-PCR analysis are listed in Table 1 Suppl. All PCR reactions were performed using *PPP Master Mix* (*Top-Bio*, Prague, Czech Republic).

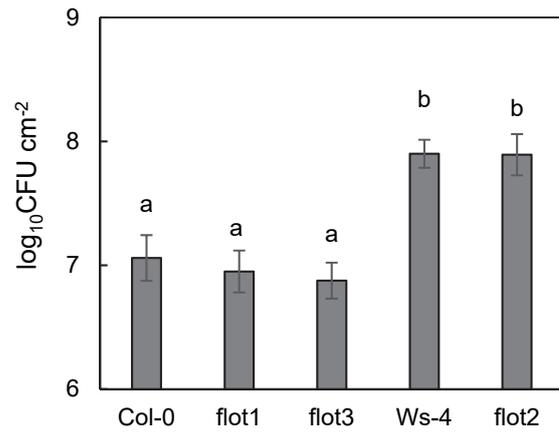


Fig. 2 Suppl. Dipping inoculation with *Pst* DC3000. Four-week-old *A. thaliana* plants were dipped in *Pst* DC3000 suspension. Values are demonstrated in log₁₀ scale of colony forming units (CFU). Means ± SEs, $n = 6$, different letters indicate statistically significant differences between the samples ($P < 0.01$, Student's *t*-test).