

Table 1 Suppl. Gene specific primers along with respective references. \* - stress inducible genes, ID – identification, Tm - melting temperature.

Genes and their abbreviations	Primer sequences (5'-3')	Gene ID	Product length [bp]	Tm [°C]	Primer efficiency [%]	Regression coefficient (R <sup>2</sup> )	Reference
Ubiquitin 2 ( <i>UBQ2</i> )	F:ATGCAAATCTTTGTGAAGACCCCTCAC R:GTGATAGTCTTGCCAGTCAAGGTCTTG	HQ340170	269	61	81.38	0.989	Brulle <i>et al.</i> (2014)
Histone 3 ( <i>H3</i> )	F:GAAATTGCACAGGACTTCAAGACTGATC R:AACCACTACTAACACTACGAACACATGCTTC	AF109910	250	59	106.47	0.965	Li <i>et al.</i> (2016)
Alpha tubulin ( <i>α-Tub</i> )	F:GTAGAACCATAACAACAGTGTCTCTCCAC R:TGGAACTCAGTGACATCCACATTGATAG	MK123512	223	59	79.93	0.962	Moraes <i>et al.</i> (2015); Reddy <i>et al.</i> (2016)
Beta tubulin ( <i>β-Tub</i> )	F:GAACCTTACAATGCCACACTATCTGTCC R:CGACCATGAAGAAATGGAGAAG	MK123513	268	59	93.07	0.995	Moraes <i>et al.</i> (2015); Reddy <i>et al.</i> (2016)
Glyceraldehyde-3-phosphate dehydrogenase ( <i>GAPDH</i> )	:CTTCTCTTCGGTGAGAAGGAGGTCAC R:CTCCTTCTCATTGACACCAACAACAAC	MK123514	213	59	80.47	0.998	Pabuayon <i>et al.</i> (2016)
Eukaryotic initiation factor-4α ( <i>eIF4-α</i> )	F:CAGCAACTTGACTATGGATTGGTGGGA R:CATCCAGCACAAACATCTTAATGTGGTC	MK123515	270	63	104.44	0.969	Gimeno <i>et al.</i> (2014)
Cyclophilin ( <i>CYP</i> )	F:CTGAGAGGATCACATTTGAGCTGTTTTG R:CACCAGTTGGTCATGCTTTAACTTG	MK123516	267	59	81.02	0.991	Pabuayon <i>et al.</i> (2016)
Actin ( <i>ACT</i> )	F:CTACCATGTTTCTGGAATTGCTGATCG R:TCAGAAGCACTTCATATGGACAATGCCAG	MK123517	249	59	105.35	0.989	Pabuayon <i>et al.</i> (2016)
Alcohol dehydrogenase ( <i>ADH</i> )*	F:GCATTCTGAGCTGTGGTATCTCTACTGG R:TTACGAAGTCAGTGACGCCAAACTTC	EU371995	216	-	-	-	Garg <i>et al.</i> (2013)
Dehydration-responsive element-binding protein 2A ( <i>DREB2A</i> )*	F:TTCTGGTTTTGCTCCTTCCACC R:ATGCTGTTTCGATTTGTGTCTTGC	KM349308.1	214	-	-	-	Sakuma <i>et al.</i> (2006)

## References

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Table 2 Suppl. Sequence information of PCR amplified genes from *Oryza coarctata* cDNA. For abbreviations of genes see Table 1 Suppl.

>*UBQ2*  
CACCATTGACAACGTCAAGGCCAAGATCCAGGACAAGGAGGGCATCCCCCGGACCAGCAGCGTCTCATCTTCGCTGGCAAGCAGCTT  
GAGGATGGCCGCACCCTGGCTGACTACACATCCAGAAGGAGTCCACCCTCCACCTTGTGCTCAGGCTCAGGGGAGGCATGCAAATCTTCG  
TCAAGACCTTGACTGGAAGAACTATCACA

>*H3*  
CTCCAGGAAGCTTGC GGAGGCATACCTTGTGGTCTCTTCGAGGACACCAACCTGTGCGCCATTCACGCCAAGCGCGTGACCATCATGCC  
TAAGGACATTGAGCTGGCCAGGAGGATTCGTGGTGAGAGGGCTTAAGTTCCCCTCGATGATTCCTTTGACAAATGAAGCATGTGTTCGTA  
GTGTTAGTAGTGGTTAA

>*α-Tub*  
CAGTTCTCCTAGACAATCGAGGCTATCTATGACATATGCCGGAGATCCCTTGACATCGAGAGGCCAACCTACACCAACTTGAACAGGCTC  
ATCTCACAGATCATATCTTCACTACCACCTCCCTGAGGTTTGATGGCGCTATCAATGTGGATGTCACTGAGTTCCAA

>*β-Tub*  
CTTGACAATGAAGCTCTATATGACATCTGCTTCCGCACTCTGAAGCTTGC GACACCCACCTTTGGTGATCTGAACCATCTTATTTCTGCAA  
CCATGATGGCGTTACCTGCTGCCTGCGCTTCCCGGGACAGCTGAATTCTGACCTCCGCAAGCTTGCAGTCAACTTGATTCCCTTCCCTCG  
TCTCCTTCTTTCTGGTCTCGA

>*GAPDH*  
GACTCCACTCATGGTCCGTTTTAAGGTTCCATTAAGGTTGTTGGATGATTCAATCCCTGGAGATCAATGGAAAGAAAGTCACAATCACTAG  
CAAAAGAGATCCTGCAGATATTCCTTGGGGTAACTTTGGAGCTGAGTATGTTGTTGAATCTTCAGGTGTA

>*eIF4-α*  
AGCTTGCCAGCAAATTGAAAAGGTCATGCGTGCTCTTGGGGACTACTTAGGTGTCAAGGTGCATGCCTGTGTTGGTGGAAACATCTGTTC  
GTGAGGACCAAAGGATTCTTGCTAGTGGCGTGCATGTTGTTGTTGGCACACCTGGACGTGTGTTTGATATGCTTCGCCGTCTATCTCTCCT  
CCATACTACATTAATCTTTGTGCTGGATGACTCCCCCCTCCCTATTCTTTT

>*CYP*  
TTCCGGGCTCTCTGCACCGGTGAGAGAGGCCTCGGAGTATCCACACAAAAGTCACTTCATTATAAGCGAACAAACATGCATCGCATTTTG  
AAAGGTTTTATGGCTCAGGGTGGTGATTTTTCAAGAGGAGATGGACGTGGTGGAGAGAGCATATATGGTGCAAAATTTGAAGATGAAAA  
CTTCAAGTTAAAGCATGCCAACCTGGTGA

>*ACT*  
AGCAGTATGAAGGTTAAAGTGATTGCACCACCAGAAAGAAAATACAGTGTCTGGATTGGTGGTTCTATTTTGGCTTCTCTGAGCACTTTC  
CAGCAGATGTGGATCTCCAAGGGCGAGTATGATGAATCTGGTCTGGCATTGTCCATATAGTGCTTCTGAA

>*Adh*  
AAAAGGCCAAGACAGTGGCTATTTTTCCGGTTTTGGAAGCTGTTGGCCTTGCTGCTATGGAAGGTGCCAGGCTGTCTGGAGCATCAGAAGG  
GAATCATTGGTGTGGACCTGAACCCTGCAAAATCGAACAAGCTAAGAAGTTTGGCTGCCTGACTTCGTAAGCATT

>*DREB2A*  
ATGCTGTTTCGATTTGTGCTTGCAATGTTTCAGCTTTGTGGAATTATTGAGTTACCTCATTGGGTCAGGA  
AGAAGAGGACGCGAAGGAAAAGCGATGGCCCTGATTCAATCACTGAAACCATCAAGTGGTGGAAAGGAGCA  
AAACCAGAA

Table 3 Suppl. Sequence validation of an amplified PCR product from *Oryza. coarctata* with *O. sativa*. For abbreviations of genes see Table 1 Suppl.

Serial number	Genes	BLASTN identity [%]	E-value
1	<i>UBQ2</i>	99	4.3e-91
2	<i>H3</i>	98	3.1e-92
3	<i><math>\alpha</math>-Tub</i>	99	9.6e-77
4	<i><math>\beta</math>-Tub</i>	99	4.3e-76
5	<i>GAPDH</i>	99	1.0e-65
6	<i>eIF4-<math>\alpha</math></i>	90	5.0e-86
7	<i>CYP</i>	99	1.2e-86
8	<i>ACT</i>	99	1.2e-70
9	<i>Adh</i>	96	1.3e-55
10	<i>DREB2A</i>	100	7.0e-76

Table 4 Suppl. Rank orders of eight selected reference genes from *Oryza coarctata* based upon their expression stability value determined by the  $\Delta Ct$  method. For abbreviations of genes see Table 1 Suppl.

Gene order	Cold		Heat		Drought		Salinity		Submergence		ABA treatment		Dual stress	
	Gene	SD												
1	<i>H3</i>	1.18	<i>ACT</i>	1.56	<i><math>\beta</math>-Tub</i>	1.16	<i>ACT</i>	1.64	<i><math>\beta</math>-Tub</i>	1.34	<i>H3</i>	1.65	<i><math>\beta</math>-Tub</i>	1.08
2	<i><math>\alpha</math>-Tub</i>	1.18	<i>CYP</i>	1.70	<i>eIF4-<math>\alpha</math></i>	1.17	<i>eIF4-<math>\alpha</math></i>	1.65	<i>ACT</i>	1.53	<i>ACT</i>	1.73	<i>eIF4-<math>\alpha</math></i>	1.21
3	<i><math>\beta</math>-Tub</i>	1.22	<i>eIF4-<math>\alpha</math></i>	1.73	<i>CYP</i>	1.19	<i>UBQ2</i>	2.03	<i>H3</i>	1.58	<i>CYP</i>	1.81	<i><math>\alpha</math>-Tub</i>	1.30
4	<i>ACT</i>	1.27	<i>UBQ2</i>	1.85	<i>ACT</i>	1.29	<i><math>\beta</math>-Tub</i>	2.16	<i>CYP</i>	1.60	<i><math>\beta</math>-Tub</i>	1.87	<i>H3</i>	1.50
5	<i>eIF4-<math>\alpha</math></i>	1.35	<i>GAPDH</i>	1.85	<i>H3</i>	1.36	<i>H3</i>	2.20	<i><math>\alpha</math>-Tub</i>	1.69	<i>GAPDH</i>	1.95	<i>ACT</i>	1.53
6	<i>GAPDH</i>	1.51	<i><math>\alpha</math>-Tub</i>	1.97	<i><math>\alpha</math>-Tub</i>	1.37	<i>GAPDH</i>	2.30	<i>GAPDH</i>	1.70	<i>eIF4-<math>\alpha</math></i>	2.14	<i>GAPDH</i>	1.66
7	<i>CYP</i>	1.60	<i><math>\beta</math>-Tub</i>	1.99	<i>UBQ2</i>	1.86	<i><math>\alpha</math>-Tub</i>	2.47	<i>eIF4-<math>\alpha</math></i>	1.77	<i>UBQ2</i>	2.46	<i>UBQ2</i>	1.69
8	<i>UBQ2</i>	1.65	<i>H3</i>	2.27	<i>GAPDH</i>	1.89	<i>CYP</i>	2.70	<i>UBQ2</i>	2.07	<i><math>\alpha</math>-Tub</i>	2.49	<i>CYP</i>	1.73

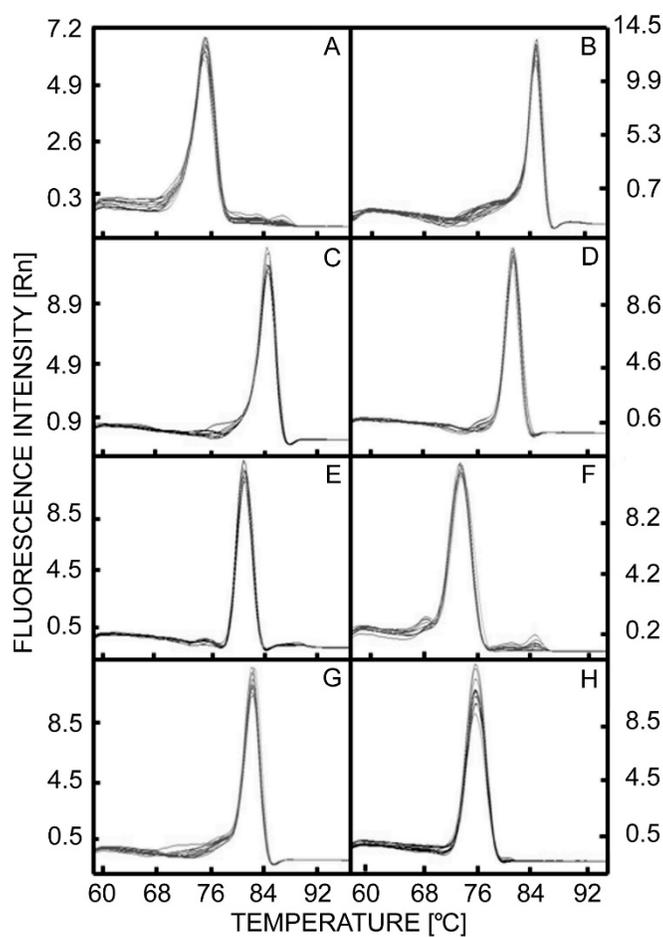


Fig. 1 Suppl. Melting curve analysis in real time quantitative PCR. Dissociation curves showing single peaks for each primer pair: *UBQ2* (A), *H3* (B),  *$\alpha$ -Tub* (C),  *$\beta$ -Tub* (D), *GAPDH* (E), *eIF4- $\alpha$*  (F), *CYP* (G), and *ACT* (H). For abbreviations of genes see Table 1 Suppl.