BRIEF COMMUNICATION

Effects of capsaicin on plant growth

H. KATO-NOGUCHI* and Y. TANAKA

Department of Biochemistry and Food Science, Faculty of Agriculture, Kagawa University, Miki, Kagawa 761-0795, Japan

Abstract

Capsaicin, a possible allelochemical, caused growth inhibition of roots and shoots of alfalfa (Medicago sativa), cress (Lepidium sativum), lettuce (Lactuca sativa), crabgrass (Digitaria sanguinalis), timothy (Phleum pratense) and ryegrass (Lolium multiflorum), and suppressed their germination. Increasing the dose of capsaicin increased the inhibition. The concentrations for 50 % inhibition of the root growth were 2.7, 0.32, 2.1, 0.27, 0.29 and 0.57 mM for alfalfa, cress, lettuce, crabgrass, timothy and ryegrass, respectively, and the concentrations for 50 % inhibition of the shoot growth were 17, 0.87, 6.7, 2.3, 1.4 and 6.2 mM for alfalfa, cress, lettuce, crabgrass, timothy and ryegrass, respectively. Germination percentage was inhibited 50 % at the concentrations 82, 88, 68, 48, 22 and 11 mM for alfalfa, cress, lettuce, crabgrass, timothy and ryegrass, respectively. Thus, effectiveness of capsaicin on the plant growth differed with species and targets, and suggests that capsaicin may act as an allelochemical to other plants.

Additional key words: alfalfa, allelopathy, Capsicum annuum, crabgrass, cress, germination inhibitor, growth inhibitor, lettuce, phytotoxicity, ryegrass, timothy.

Capsaicin, a primary pungent principle contained in a variety of Capsicum spp. such as chili pepper, red pepper and cayenne pepper, is an amide derivative of vanillylamine and 8-methyl-4-nonal-6-enolic acid (Ochoa-Alejo and Salgado-Garacigila 1992). Capsaicin has a number of pharmacological and physiological effects on mammals (Suh and Lee 1995, Abdel-Salam et al. 1997), however, the physiological role of capsaicin in plants is not clear and only limited information is available in the literature with regard to its effectiveness on plant growth (Cho et al. 1992). Thus, in the present research, the effects of capsaicin on germination and growth of six plant species were determined.

Capsaicin, obtained from Nacalai Chemicals (Kyoto, Japan), was dissolved in methanol, added to a sheet of filter paper (No. 2, Toyo Ltd. Tokyo, Japan) in a 5.5-cm Petri dish, and dried. After placement of the methanol, the filter paper in Petri dish was moistened with 2 cm² of 0.05 % (v/v) aqueous Tween 20. The concentrations of capsaicin in the bioassay were 0, 0.01, 0.03, 0.1, 0.3, 1, 3, 10, 30, 100 mM. Fifty seeds of alfalfa (Medicago sativa L.), cress (Lepidium sativum L.), lettuce (Lactuca sativa L.), crabgrass (Digitaria sanguinalis L.), timothy (Phleum pratense L.), and ryegrass (Lolium multiflorum Lam.) were separately arranged on the filter paper and allowed to germinate in the dark at 25 °C for 24 h (alfalfa, cress, lettuce) or 60 h (crabgrass, ryegrass, timothy). Then, the germination percentage was calculated by reference to that of control seeds treated with plain solution (Kato-Noguchi 2001).

For further experiment, seeds of alfalfa, cress, crabgrass, lettuce, timothy and ryegrass were sown on the filter paper and allowed to germinate in the dark at 25 °C for 12 h (alfalfa, cress, lettuce) or 48 h (ryegrass, timothy, crabgrass). Capsaicin was dissolved and added to a sheet of filter paper in 3.5-cm Petri dish and the filter paper was moistened with 0.8 cm² of 0.05 % (v/v) aqueous Tween 20 as described above. Then, 10 germinated seeds of test plants were arranged on the filter paper and allowed to grow in the dark at 25 °C for 48 h. The shoot and root lengths of the seedlings were measured, and the percentage length of the seedlings was calculated by

Received 10 June 2002, accepted 4 September 2002.
Abbreviations: IC50 - concentration required for 50 % inhibition in the assay.
* Corresponding author; fax: (+81) 87 8913086, e-mail: hisashi@ag.kagawa-u.ac.jp

157
reference to that of the control plants treated with plain solution. All experiments were repeated three times in completely randomized block designs.

Capsaicin suppressed germination of lettuce seeds, and inhibited growth of their roots and shoots at concentrations greater than 3, 0.1 and 0.3 mM, respectively, with the most marked inhibition being achieved on the root growth (Fig. 1). When the percentage inhibition was plotted against the logarithm of the concentrations of capsaicin, the response curves of the germination, root growth and shoot growth were linear between 10 and 60 %, 10 and 80 % and 10 and 85 % inhibition, respectively. The effectiveness on the germination was weak and complete response curve was not obtained. The concentrations required for 50 % inhibition in the assay (defined as I50) were 68, 2.1 and 6.7 mM for the germination, root growth and shoot growth, respectively, as interpolated from the response curves. Comparing I50 values, the inhibitory effect of capsaicin on the root and shoot growth was 26- and 10-fold greater than that on the germination, respectively.

The effects of capsaicin on root and shoot growth, and germination of all test species were examined and I50 values were determined from the concentration-response curves as described above (Table 1). Capsaicin concentration-dependently inhibited the germination, root and shoot growth of all species although its effectiveness differed with species and targets. In all bioassays, the I50 values of the root growth were smallest, followed in order by the shoot growth and the germination, confirming that the root growth was the most inhibited. Additionally, capsaicin was much more effective on the germination of monocotyledonous species (crabgrass, timothy and ryegrass) than that of dicotyledonous species (alfalfa, cress and lettuce). The difference between dicotyledonous and monocotyledonous species was less clear in shoot and root growth than in germination percentage.

It was found that many secondary metabolite are released into the environment, either as exudation from living plant tissues or by decomposition of plant material under certain conditions (Rice 1984, Putnam 1988, Einhellig 1996). These findings together with the occurrence of capsaicin in Capsicum plants (Leece and Louden 1968, Iwai et al. 1979) and its effectiveness on growth (Fig. 1 and Table 1) suggest that capsaicin may act as an allelochemical of pepper to neighboring or successional plants after being released into the environment by the decomposition of the plants in the soil or by the exudation from their roots.

Controlling weeds through allelopathy is one strategy to reduce synthetic chemical herbicide dependency in the present weed management systems (Rice 1984, Putnam 1988). It has also been shown that certain plant residues and extracts may function as weed suppressive agents (Putnam 1988, Einhellig 1996, Kato-Noguchi 2001, Kato-Noguchi and Ino 2001). Thus, it is possible that the pepper itself may be important as a weed suppressive agent.

Table 1. I50 values [mM] of capsaicin for root and shoot growth, and germination of test plants. Means ± SE from three replicates (10 plants each for determination of root and shoot growth, and 50 seeds each for determination of germination).

<table>
<thead>
<tr>
<th>Species</th>
<th>Germination</th>
<th>Root</th>
<th>Shoot</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alfalfa</td>
<td>82.0 ± 5.7</td>
<td>2.70 ± 0.15</td>
<td>17.00 ± 1.10</td>
</tr>
<tr>
<td>Cress</td>
<td>88.0 ± 6.8</td>
<td>0.32 ± 0.02</td>
<td>0.87 ± 0.05</td>
</tr>
<tr>
<td>Lettuce</td>
<td>68.0 ± 5.9</td>
<td>2.10 ± 0.13</td>
<td>6.70 ± 0.41</td>
</tr>
<tr>
<td>Crabgrass</td>
<td>4.8 ± 0.2</td>
<td>0.27 ± 0.01</td>
<td>2.30 ± 0.17</td>
</tr>
<tr>
<td>Timothy</td>
<td>22.0 ± 1.8</td>
<td>0.29 ± 0.01</td>
<td>1.40 ± 0.01</td>
</tr>
<tr>
<td>Ryegrass</td>
<td>11.0 ± 0.9</td>
<td>0.57 ± 0.03</td>
<td>6.20 ± 0.39</td>
</tr>
</tbody>
</table>

Fig. 1. Effects of capsaicin on root and shoot growth, and germination of lettuce. Means ± S.E. from three replicates (50 seeds each for determination of germination, and 10 plants each for determination of root and shoot growth). Germination rate of control plants was 91 ± 7.3 %, and length of control plants was 18.6 ± 1.1 and 6.9 ± 0.33 mm for roots and shoots, respectively.

References
